GABAergic Drugs can Enhance or Attenuate Chlordiazepoxide-Induced Sleep Time in a Heterogeneous Strain of Mice

TODD D. MCINTYRE AND HERBERT P. ALPERN*1

*Behavioral Neuroscience Program, Department of Psychology, University of Colorado, Boulder, CO 80309

Received 10 March 1986

McINTYRE, T. D. AND H. P. ALPERN. GABAergic drugs can enhance or attenuate chlordiazepoxide-induced sleep time in a heterogeneous strain of mice. PHARMACOL BIOCHEM BEHAV 25(5) 1077–1081, 1986.—Evidence supports the notion that differences between the Long-Sleep and Short-Sleep selectively-bred lines of mice are attributable to differences in brain excitability and that these differences are mediated by activity of the GABAergic system. The general applicability of this hypothesis to other populations of mice was tested by using an outbred strain of mice. Specifically, a heterogeneous strain of mice was administered several doses of the hypotic chlordiazepoxide. Additionally, the indirect GABA agonist AOAA, and the GABA antagonists bicuculline, picrotoxin and pentyleneterazol were administered to independent groups in conjunction with chlordiazepoxide. The results clearly demonstrate that chlordiazepoxide dosedependently increased hyponsis, while AOAA enhanced, and the antagonists attenuated sleep time. These findings can be used to support the contention that GABA mediates the bidirectional response of Long-Sleep and Short-Sleep mice to CNS hypnotic-depressants; and, further, show that GABA mediation of sleep time in mice is a general phenomenon.

Heterogeneous mice	Pharmacogenetics	GABA	AOAA	Bicuculline	Picrotoxin	Pentylenetetrazol
--------------------	------------------	------	------	-------------	------------	-------------------

A useful strategy for establishing animal models (i.e., particular phenotypes) is the selective breeding method [23,24]. Typically, selective breeding involves the testing of numerous animals for some phenotype of interest, and the subsequent mating of those animals of high phenotypic value with others of like phenotypic value, and similarly for animals of low phenotypic value. Unlike inbreeding, though, sib and cousin matings are avoided. Then, as selection pressure is maintained, the offspring of each succeeding generation will exhibit a corresponding increase in the number of "high" and "low" alleles at the pertinent loci. This increase in the number of high and low alleles is a function of the degree to which the particular phenotype is heritable [24]. Most importantly, because inbreeding is proscribed, all other alleles not influencing the selected trait should assort randomly.

One such selection study, which was initiated to establish heritability of factors contributing to alcohol sensitivity, was begun by McClearn and colleagues [22,24]. Briefly, two lines of mice that differ bidirectionally in their sensitivity to a hypnotic dose of ethanol were derived from a genetically heterogeneous strain (HS). The HS was created by systematically crossing eight inbred strains of mice (A, AkR, BALB/c, C3H/2, C57/BL, DBA, Is/Bi, and RIII) [21]. Animals from this population were examined with respect to loss of righting reflex (sleep time) subsequent to an intraperitoneal injection of ethanol. Males and females that exhibited long sleep times were mated, as were males and females exhibiting minimal or no loss of the righting reflex. After 16 generations, no overlap in the distributions of sleep time was discernible. Those demonstrating extended sleep times were designated the Long-Sleep (LS) line, while those that did not lose the righting reflex, or that lost it for only a brief period, were termed the Short-Sleep (SS) line.

Initially it was proposed that these lines were selected for specific sensitivity to alcohol [10, 13, 14, 31], but recent evidence shows that this is not the case [1, 26, 27], and we have proposed a unifying hypothesis which accounts for the many differences between the LS and SS selected lines [26]. Since the LS line is more sensitive than the SS line to the soporific effects of alcohols [10,14], barbiturates [1, 2, 25, 27], benzodiazepines [27], and miscellaneous hypnotics (e.g., adenosine and nitrous oxide; [9,17]), it seemed reasonable to conclude that these lines were originally selected for differences in tonic brain excitability, and that these differences are related to activity at the GABA synapse.

If our hypothesis concerning GABAergic mediation of differences between LS and SS selected lines is generally applicable, then drug effects observable in the LS and SS lines should also be found in non-selected outbred animals such as the HS strain. Since the HS strain was maintained during the selection program as a control for evaluating random intergenerational variability, they are ideal for analyzing the relationship between the original operational selection criterion (ethanol-induced hypnosis) and whatever CNS mechanism is mediating the bidirectional response.

There is a growing body of evidence implicating the

Requests for reprints should be addressed to H. P. Alpern.

1078

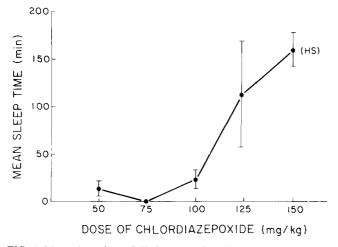


FIG. 1. Mean sleep time±S.E. for each of the independent groups of HS mice administered chlordiazepoxide.

GABAergic system in the mediation of the hypnotic effects of most CNS depressants [6, 7, 29, 33, 34, 37, 39, 43], and the evidence for benzodiazepines is the most convincing. Consequently, this paper reports experiments that evaluate chlordiazepoxide-induced sleep time, as well as the effects of GABAergic agents on induced sleep time in HS mice. Since the effects of chlordiazepoxide on the LS and SS lines has already been established, it was selected to test the generality of our hypothesis. Initially a dose-response function was constructed, and then the effect of the indirect GABA agonist amino-oxyacetic acid (AOAA) on chlordiazepoxideinduced sleep time was determined. It was our prediction that this GABA agonist, which blocks the GABA catabolic enzyme GABA-transaminase, would enhance sleep time in these mice. To the contrary, we speculated that GABA antagonists would attenuate chlordiazepoxide-induced hypnosis. To test this, we first evaluated the effect that two doses of the classical GABA antagonist bicuculline had on induced sleep time. Since bicuculline is a direct GABA receptor antagonist [32], and since benzodiazepines bind at different locations on the GABA receptor complex [39], we also employed two analeptics (picrotoxin and pentylenetetrazol) which exert their convulsive effects via an allosteric subsite on the benzodiazepine-GABA receptor-chloride ionophore [32,42].

Animals

METHOD

One hundred and four HS mice 100–150 days old served as subjects. These mice were descendants of animals from the 39th production generation, obtained from the Institute for Behavioral Genetics, Boulder, CO 80309. Littermates were housed together, but randomly distributed across experimental groups. All animals were maintained on a 12 hr light/dark cycle with food and water continuously available. Each independent group in the four experiments consisted of eight males, except the final experiment, where each group consisted of four males and four females.

Procedure

Each group in Experiment 1 received either 50, 75, 100,

ΤA	BL	Æ	1	
----	----	---	---	--

MEAN SLEEP TIME (MIN) ± S.E. FOR THE CONTROL GROUP
(0 mg/kg AOAA) AND THE GROUPS TREATED WITH
10 or 20 mg/kg OF AOAA 2 HR PRIOR TO 100 mg/kg
CHLORDIAZEPOXIDE (N=8 FOR EACH GROUP)

AOAA (mg/kg, IP)	Sleep Time	Significance (when compared with 0 mg/kg group)
0.0	22.7 ± 9.5	
10.0	16.7 ± 8.3	ns
20.0	182.6 ± 65.2	p<0.01

125, or 150 mg/kg chlordiazepoxide (Librium) in an injection volume of 0.1 ml/g body weight. For Experiment 2, each group received either 0.0, 10.0, or 20.0 mg/kg aminooxyacetic acid hemihydrochloride, followed 2 hr later by 100 mg/kg of chlordiazepoxide. Each group in Experiment 3 received 150 mg/kg chlordiazepoxide. Then 15 min after sleep time began, groups were administered either saline, 1.0 or 2.0 mg/kg (+)-bicuculline dissolved in saline adjusted to a pH of 2.5. For Experiment 4, each group received 150 mg/kg chlordiazepoxide, and then 15 min later, either saline, 7.0 mg/kg picrotoxin or 50 mg/kg pentylenetetrazol. All injections were made intraperitoneally and it should be noted that with this route of administration the doses of bicuculline, picrotoxin and pentylenetetrazol selected were subconvulsive. Sleep time was assessed in the following manner. After injection of chlordiazepoxide, an animal was placed on its back in a V-shaped (90° angle) Plexiglas sleep trough until it was unable to right itself four times within sixty seconds, at which time it was considered to have lost its righting reflex. An animal was considered to have regained its righting reflex when it could right itself four times within a sixty second period.

RESULTS

Figure 1 displays the dose-response function for chlordiazepoxide. As can be seen, the animals began sleeping to a significant degree at 100 mg/kg, and the animals sleept progressively longer at the two higher doses (125 and 150 mg/kg). A oneway analysis of variance was performed on the three highest doses, which constitute the positively accelerating portion of the dose-response function. The analysis was significant F(2,21)=4.04, p<0.05, and group comparisons using the protected *t*-test indicate that only the 100 and 150 mg/kg groups are significantly different from each other (p<0.05).

As can be seen in Table 1, which displays the results of administering different doses of AOAA prior to chlordiazepoxide, the highest dose of AOAA greatly enhanced sleep time. A one-way analysis of variance was used to analyze these data, F(2,21)=6.01, p<0.01, and group comparisons using the protected *t*-test indicate that only the 20 mg/kg dose is significantly different (p<0.01) from the control dose (0 mg/kg).

Т	A	в	L	Æ	2

MEAN SLEEP TIME (MIN) ± S.E. FOR THE GROUPS RECEIVING 150 mg/kg CHLORDIAZEPOXIDE AND THEN 15 MIN AFTER HYPNOSIS BEGAN, EITHER SALINE OR 1.0 OR 2.0 mg/kg OF BICUCULLINE HYDROCHLORIDE (N=8 FOR EACH GROUP)

Bicuculline (mg/kg, IP)	Sleep Time	Significance (when compared with 0 mg/kg group)
0.0	140.5 ± 13.5	
1.0	62.7 ± 18.2	p<0.01
2.0	26.6 ± 2.3	p < 0.01

MEAN SLEEP TIME (MIN) ± S.E. FOR THE GROUPS RECEIVING 150 mg/kg CHLORDIAZEPOXIDE AND THEN 15 MIN AFTER HYPNOSIS BEGAN, EITHER SALINE OR 7.0 mg/kg PICROTOXIN OR 50 mg/kg PENTYLENETETRAZOL (N=8 FOR EACH GROUP)

Treatment	Sleep Time	Significance (when compared with 0 mg/kg group)	
Saline	195.0 ± 17.2		
Picrotoxin	33.0 ± 1.6	p < 0.01	
Pentylenetetrazol	23.1 ± 1.6	p < 0.01	

Table 2 illustrates the results of administering different doses of bicuculline after the induction of chlordiazepoxide sleep time. As can be seen, both doses of bicuculline attenuated sleep time, and a one-way analysis of variance used to analyze these data is highly significant, F(2,21)=19.59, p<0.01. Group comparisons made with the protected *t*-test confirm that both groups administered bicuculline are significantly different than the saline control group (p<0.01).

Finally, Table 3 displays the results of administering either picrotoxin or pentylenetetrazol subsequent to the induction of chlordiazepoxide hypnosis. As can be readily seen, both the these analeptics also significantly attenuated sleep time. This observation was verified with a one-way analysis of variance, F(2,21)=92.85, p<0.01, and protected *t*-tests confirm that each analeptic significantly attenuated chlordiazepoxide-induced sleep time (p<0.01).

DISCUSSION

The results of these experiments indicate that nonselected outbred mice (HS) react to GABA drugs in a manner that is similar to the LS and SS lines. We have previously demonstrated that the LS and SS lines exhibit differential hypnotic reactions to the anxiolytic chlordiazepoxide, and further, that the indirect GABA agonist AOAA dosedependently increased the induced sleep time [25, 27]. These results, therefore, support the generality of a GABA hypothesis for the mediation of sleep time in mice. One result obtained deserves further comment. In the second experiment the 10 mg/kg dose of AOAA did not affect induced sleep time. This dose was apparently too low and it should be mentioned that the AOAA doses were selected on the basis of previous work with LS and SS mice, where it was found that doses higher than 20 mg/kg did not further enhance sleep time for LS mice. A final comment concerning experimental design should be made. The final experiment employed only one dose of picrotoxin and pentylenetetrazol since previous work had shown that these doses produce effects on convulsive threshold that are similar to 2 mg/kg of bicuculline. As noted earlier these doses are subconvulsive and the clonus ED₂₅s for the three drugs are: bicuculline, 4 mg/kg; picrotoxin, 10 mg/kg; pentylenetetrazol, 65 mg/kg.

As previously outlined, the LS and SS display a bidirectional response to alcohols, barbiturates, benzodiazepines and adenosine, all of which have had GABA transmission implicated in their mode of action. More direct evidence also exists. For instance, the GABA agonists THIP and baclofen were significantly more effective in inhibiting bar-holding in LS mice than they were in SS mice [19]. Further, in a gridtest of locomotor coordination, several alcohols and pentobarbital all produce greater coordination difficulties in LS mice than in SS mice [8]. It has been demonstrated recently that intraventricularly administered picrotoxin dramatically reduced ethanol-induced sleep time in both lines [20], and it was concluded that this demonstrates a non-specific excitatory effect and is not necessarily linked to GABA. The argument is that if a specific neurochemical deficit exists in LS and SS mice, then correcting such a deficit with an agent like picrotoxin should produce decreased sleep time for LS mice, but increased sleep time for SS mice. This appealing explanation overlooks the fact that GABA works by increasing chloride conductance, while various GABA blockersanaleptics block this increase in conductance [32,43]. It is not apparent how a particular drug would decrease chloride conductance in one selected line, while increasing this conductance in the other line.

Benzodiazepines enhance primary afferent depolarization [5] in a bicuculline-sensitive fashion [35]. Further, benzodiazepines antagonize the increase in cerebellar cGMP caused by depletion of GABA [7]. Rapid, reversible, saturable, and stereospecific binding of [3H]-diazepam has been demonstrated in brain [3, 40, 41], while high correlations between the ability of benzodiazepines to displace diazepam and their efficacy in several behavioral tests has also been reported [41]. It may be more appropriate to envision "benzodiazepine receptors" as recognition sites for which a wide variety of agents have an affinity (i.e., benzodiazepines, triazolopyradazines, cyclopyrolones, quinolines, β -carbolines, and pyrazologuinolinones). This finding has engendered a classification of ligands as either full agonists, partial agonists, inverse agonists or antagonists [4, 11, 12, 15, 28, 32, 34, 37–39]. Finally, in addition to peripheral binding sites [36], evidence exists for two high-affinity neuronal sites termed Type I and Type II [16,18], which have a heterogeneous CNS distribution [44].

Overall, the evidence reported here supports our original contention that GABA mediates the bidirectional reactions of LS and SS mice to CNS hypnotic-depressants and shows that GABA mediation of depressant-induced sleep time is applicable to mice in general. Several rather complex models of the benzodiazepine-GABA receptor-chloride ionophore have been proposed [12, 28, 30, 32, 34, 37, 39], and one of these may eventually be useful in elucidating the mechanisms responsible for the behavioral results that have been reported with hypnotic-depressants.

MCINTYRE AND ALPERN

REFERENCES

- Alpern, H. P. and T. D. McIntyre. Evidence that the selectively-bred Long- and Short-Sleep mouse lines display common narcotic reactions to many depressants. *Psychophar*macology (Berlin) 85: 456–459, 1985.
- Alpern, H. P. and T. D. McIntyre. Sedative-hypnotic anomalies related to dose of pentobarbital in Long-Sleep and Short-Sleep selectively-bred mice. *Pharmacol Biochem Behav* 25: 333–336, 1986.
- 3. Braestrup, C. and R. F. Squires. Brain specific benzodiazepine receptors. Br J Psychiatry 133: 249–260, 1978.
- Braestrup, C., T. Honore, M. Nielsen, E. R. Petersen and L. H. Jensen. In: *Benzodiazepine Recognition Site Ligands: Biochemistry and Pharmacology*, edited by G. Biggio and E. Costa. New York: Raven, 1983, pp. 29–36.
- Chin, J. H., D. P. Crankshaw and J. J. Kendig. Changes in the dorsal root potential with diazepam and with the analgesics aspirin, nitrous oxide, morphine, and meperidine. *Neuropharmacology* 13: 305-315, 1974.
- Costa, E., A. Guidotti, C. C. Mao and A. Suria. New concepts on the mechanism of action of benzodiazepines. *Life Sci* 17: 167–186, 1975.
- Costa, E. and A. Guidotti. Molecular mechanisms in the receptor action of benzodiazepines. *Annu Rev Pharmacol Toxicol* 19: 531-545, 1979.
- Dudek, B. C. and T. J. Phillips. Locomotor stimulant and intoxicant properties of methanol, ethanol, tertiary butanol and pentobarbital in Long-Sleep and Short-Sleep mice. *Subst Alcohol Actions Misuse* 4: 31-36, 1983.
- Dunwiddie, T. V. and W. R. Proctor. Behavioral sensitivity to purinergic drugs parallels ethanol sensitivity in selectively bred mice. *Science* 217: 519–521, 1984.
- Erwin, V. G., W. D. W. Heston, G. E. McClearn and R. A. Deitrich. Effect of hypnotics on mice genetically selected for sensitivity to alcohol. *Pharmacol Biochem Behav* 4: 679–683, 1976.
- Gallagher, D. W., P. Mallorga, W. Oertel, R. Henneberry and J. F. Tallman. ^aH-diazepam binding in a mammalian central nervous system: A pharmacological characterization. *J Neurosci* 1: 218–225, 1981.
- Haefely, W. Antagonists of benzodiazepines: Functional aspects. In: *Benzodiazepine Recognition Site Ligands: Biochemistry and Pharmacology*, edited by G. Biggio and E. Costa. New York: Raven Press, 1983, pp. 73-94.
 Howerton, T. C., M. F. O'Connor and A. C. Collins. Lipid
- Howerton, T. C., M. F. O'Connor and A. C. Collins. Lipid solubility may control the hypnotic and hypothermic response of Long-Sleep and Short-Sleep mice to alcohols, barbiturates, methprylon, and ethchlorvynol. *Alcoholism: Clin Exp Res* 6: 74, 1982 (Abstract).
- Howerton, T. C., M. E. O'Connor and A. C. Collins. Differential effects of long-chain alcohols in long- and short-sleep mice. *Psychopharmacology (Berlin)* **79**: 313–317, 1983.
- Hunkeler, W., H. Mohler, L. Pieri, P. Polc, E. P. Bonetti, R. Cumin, R. Schafener and W. Haefely. Selective antagonists of benzodiazepines. *Nature* 290: 514–516, 1981.
- Klepner, C. A., A. A. Lippa, D. I. Benson, M. C. Sano and B. Beer. Resolution of two biochemically and pharmacologically distinct benzodiazepine receptors. *Pharmacol Biochem Behav* 11: 457–462, 1979.
- Koblin, D. D. and J. E. Deady. Anaesthetic requirement in mice selectively bred for differences in ethanol sensitivity. *Br J Anaesth* 53: 5-10, 1981.
- Lippa, A., C. Klepner, D. Benson, D. Critchett, M. Sano and B. Beer. The role of GABA in mediating the anticonvulsant properties of benzodiazepines. *Brain Res Bull* 5: 861–865, 1980.
- Martz, A., R. A. Deitrich and R. A. Harris. Behavioral evidence for the involvement of γ-aminobutyric acid in the actions of ethanol. *Eur J Pharmacol* 89: 53-62, 1983.

- Masserano, J. M. and N. Weiner. Investigations into the neurochemical mechanisms mediating differences in ethanol sensitivity in two lines of mice. *J Pharmacol Exp Ther* 221: 404–409, 1982.
- McClearn, G. E., J. R. Wilson and W. Meredith. The use of isogenic and heterogenic mouse stocks in behavioral research. In: Contributions to Behavior-Genetic Analysis: The Mouse as a Prototype, edited by G. Lindzey and D. D. Thiessen. New York: Appleton-Century-Crofts, 1970, pp. 3-22.
- 22. McClearn, G. E. and R. Kakihana. Selective breeding for ethanol sensitivity in mice. *Behav Genet* **3**: 409–410, 1973.
- McClearn, G. E. Animal models of genetic factors in alcoholism. In: *Advances in Substance Abuse*, vol 11, edited by N. K. Mello. Greenwich, CT: Jai Press, 1981, pp. 25–51.
- 24. McClearn, G. E. and R. Kakihana. Selective breeding for ethanol sensitivity: Short-sleep and long-sleep mice. In: *Development of Animal Models as Pharmacogenetic Tools*. (DHHS Publication No. 81-1133), edited by G. E. McClearn, R. A. Deitrich and V. G. Erwin. Washington, DC: U. S. Government Printing Office, 1981, pp. 147–159.
- McIntyre, T. D. and H. P. Alpern. The interaction of CNS depressants and GABAergic drugs in mice selectively-bred for the narcotic effects of ethanol. Soc Neurosci Abstr 9: 130, 1983.
- McIntyre, T. D. and H. P. Alpern. Reinterpretation of the literature indicates differential sensitivities of Long-Sleep and Short-Sleep mice are not specific to alcohol. *Psychopharmacol*ogy (*Berlin*) 87: 379–389, 1985.
- McIntyre, T. D. and H. P. Alpern. Thiopental, phenobarbital, and chlordiazepoxide induce the same differences in narcotic reaction as ethanol in Long-Sleep and Short-Sleep selectivelybred mice. *Pharmacol Biochem Behav* 24: 895–898, 1986.
- Mohler, H. Benzodiazepine receptors: Differential ligand interactions and purification of the receptor protein. In: *Benzodiazepine Recognition Site Ligands: Biochemistry and Pharmacology*, edited by G. Biggio and E. Costa, New York: Raven, 1983, pp. 47-56.
- 29. Nestoros, J. N. Anxiety as a state of diminished GABAergic neurotransmission resulting from too frequent recruitment of GABAergic neurons: A neurophysiological model. *Prog Neuropsychopharmacol* 5: 591–594, 1981.
- Nielsen, M., T. Honore and C. Braestrup. Radiation inactivation of brain [³⁵S]-butylbicyclophosphorothionate binding sites reveals complicated molecular arrangements of the GABA/ benzodiazepine receptor chloride channel complex. *Biochem Pharmacol* 34: 3633-3642, 1985.
- O'Connor, M. F., T. C. Howerton and A. C. Collins. Effects of pentobarbital in mice selected for differential sensitivity to ethanol. *Pharmacol Biochem Behav* 17: 245–248, 1982.
- 32. Olsen, R. W., M. K. Ticku, D. Greenlee and P. VanNess. GABA receptor and ionophore binding sites: Interactions with various drugs. In: GABA Neurotransmitters. edited by P. Krogsgaard-Larsen, J. Scheel-Kruger and H. Kofod. Munksgaard: Copenhagen, 1979.
- Olsen, R. W. Drug interactions at the GABA receptorionophore complex. Annu Rev Pharmacol Toxicol 22: 245–277, 1982.
- Paul, S., P. Marangos and P. Skolnick. The benzodiazepine-GABA-chloride ionophore receptor complex: Common site of minor tranquilizer action. *Biol Psychiatry* 16: 213–229, 1981.
- Pole, P., H. Mohler and W. Haefely. The effect of diazepam on spinal cord activities: Possible sites and mechanisms of action. *Naunyn Schmiedebergs Arch Pharmacol* 284: 319–327, 1974.
- 36. Schoemaker, H., R. G. Boles, W. D. Horst and H. I. Yamamura. Specific high affinity binding sites for [³H]RO 5-4864 in rat brain and kidney. *J Pharmacol Exp Ther* 225: 61–69, 1982.
- 37. Skolnick, P. and S. Paul. Benzodiazepine receptors in the central nervous system. *Int Rev Neurobiol* 23: 103–106, 1982.

- Skolnick, P. and S. M. Paul. Benzodiazepines and nonbenzodiazepines. In: *Receptor Binding in Drug Research*, edited by R. Obrien. New York: Marcel Dekker, in press, 1986.
- 39. Skolnick, P., H. Havoundjian and S. M. Paul. Modulation of the benzodiazepine-GABA receptor complex by multiple allosteric sites: Evidence for a barbiturate receptor. In: *Clinical Pharmacology in Psychiatry*, edited by S. Dahl, L. Gram, S. Paul and W. Putter. Berlin: Springer-Verlag, 1986.
- 40. Squires, R. F. and C. Braestrup. Benzodiazepine receptors in rat brain. *Nature* 266: 732-734, 1977.
- 41. Tallman, J. F. and D. W. Gallagher. The GABAergic system: A locus of benzodiazepine action. *Annu Rev Neurosci* 8: 21–41, 1985.
- Ticku, M. K., M. Ban and R. W. Olsen. Binding of ³Hdihydropicrotoxinin, a gamma aminobutyric acid synaptic antagonist to rat brain membranes. *Mol Pharmacol* 14: 391–402, 1978.
- 43. Ticku, M. K. and W. C. Davis. Evidence that ethanol and pentobarbital enhance [³H]-diazepam binding at the benzodiazepine-GABA receptor-ionophore complex indirectly. *Eur J Pharmacol* 71: 521-522, 1981.
- 44. Young, W. S., D. Niehoff, N. J. Kunar, B. Beer and A. Lippa. Multiple benzodiazepine receptor localization by light microscopic radiohistochemistry. J Pharmacol Exp. Ther 216: 425– 430, 1981.

,